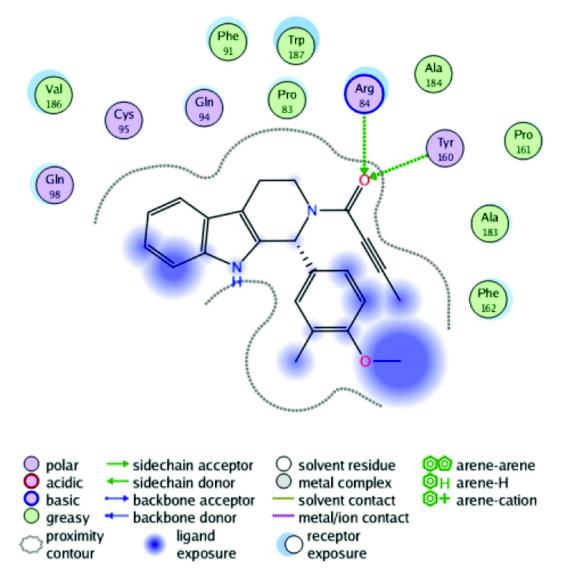
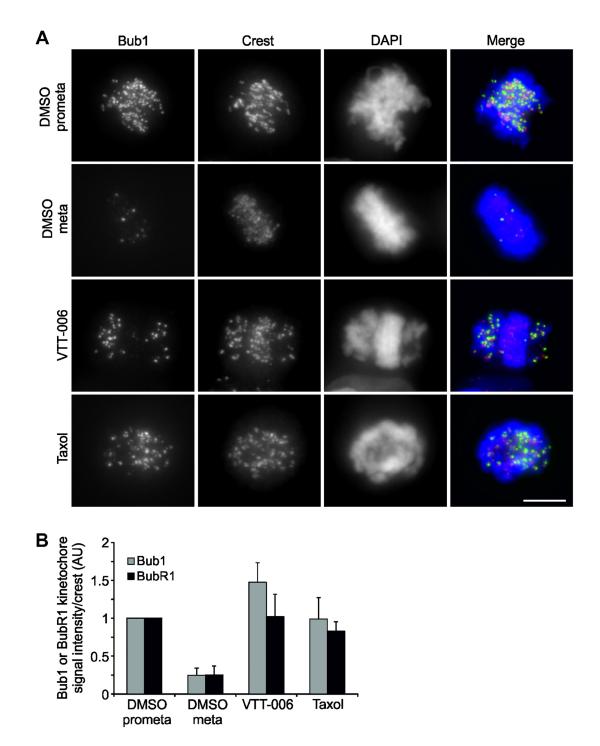
VTT-006, an anti-mitotic compound, binds to the Ndc80 complex and suppresses cancer cell growth *in vitro*

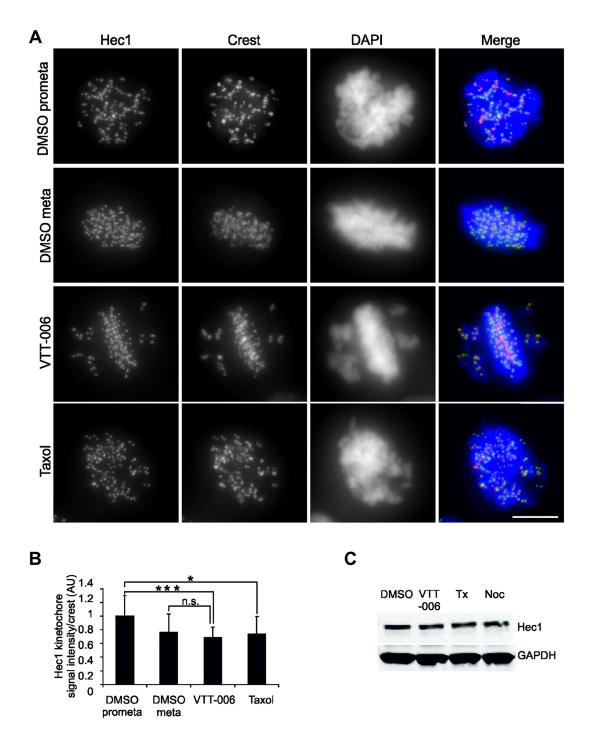
SUPPLEMENTARY MATERIAL



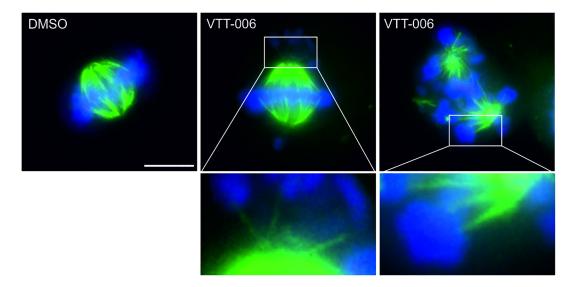
Supplementary Figure 1: Predicted interactions between VTT-006 and Hec1 CH domain. Various amino acid residues located in the Hec1 CH domain are shown. VTT-006 is predicted to interact via H-bonds between carbonyl oxygen of VTT-006 and both Arg84 and Tyr160 of Hec1. In addition, the structure of VTT-006 is proposed to have favourable vdW-interactions with the lipophilic residues of the binding cavity. Image was created with Molecular Operating Environment (MOE).



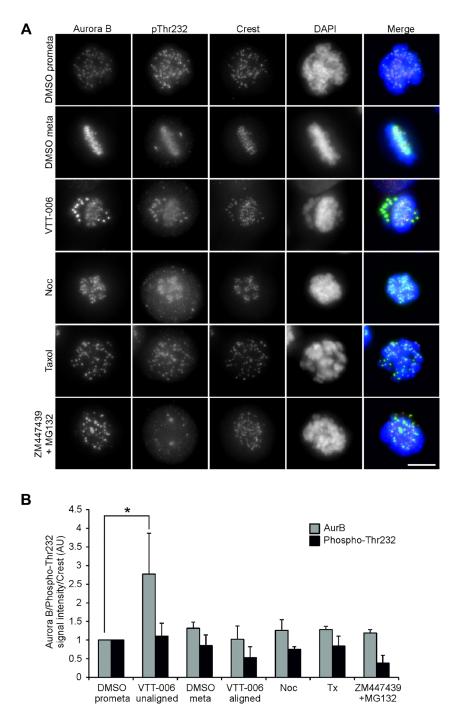
Supplementary Figure 2: Mitotic checkpoint is active in VTT-006 treated cells. HeLa cells were treated with DMSO, 10 μ M VTT-006 or 100 nM Taxol for 6 h, fixed and stained with anti-Bub1, anti-BubR1 and crest antibodies. (A) Images from Bub1 staining showing clear signals at the kinetochores of unaligned chromosomes in VTT-006 treated cells indicating continued spindle checkpoint activity. Merge shows Bub1 (green), crest (red) and DAPI (blue). (B) Quantification of Bub1 and BubR1 kinetochore signals. For VTT-006, only unaligned chromosomes near spindle poles were quantified. Result is average \pm SD from 3 replicate experiments (8–15 cells and 20 kinetochores per cell were quantified in each experiment). Scale bar = 10 μ m.



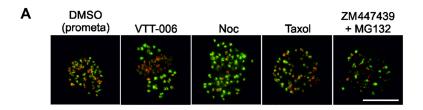
Supplementary Figure 3: Analysis of Hec1 kinetochore level and total protein level in VTT-006 treated cells. (A) Images of HeLa cells stained with Hec1 antibody (green in merge) and crest (red). (B) Quantification of Hec1 kinetochore level. Average of 15 cells (300 randomly selected individual kinetochores) \pm SD is shown. (C) Western blot showing Hec1 and GAPDH after treatment with indicated compounds. HeLa cells were treated with DMSO, 10 μ M VTT-006 or 100 nM Taxol for 12 h before fixing for immunofluorescence or collecting cells for Western blot. Scale bar = 10 μ m.

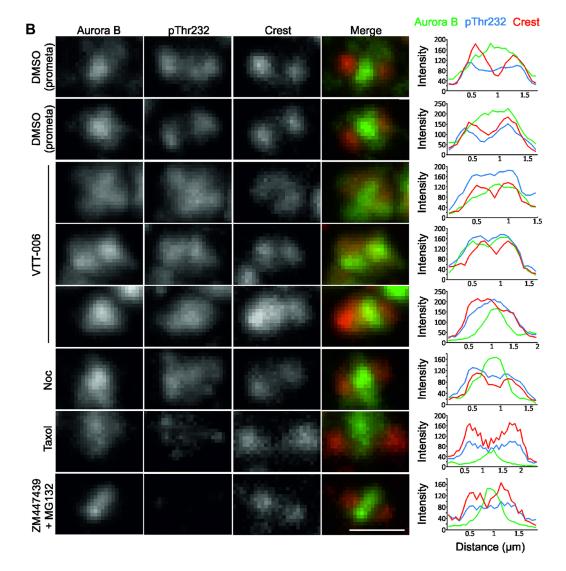


Supplementary Figure 4: Kinetochore-microtubule attachments are stable in VTT-006 treated cells. Overlays showing Hela cells treated with DMSO or 5 μ M VTT-006 for 4 h, lysed in cold Ca ⁺⁺ buffer and stained for tubulin (green) and DNA (blue). Insets show high magnification views of microtubules extending towards unaligned chromosomes. Scale bar = 10 μ m.



Supplementary Figure 5: VTT-006 increases Aurora B accumulation at centromeres of unaligned chromosomes. (A) Immunofluorescent images of cells treated with indicated compounds showing Aurora B accumulation at centromeres of unaligned chromosomes. (B) Quantification of Aurora B and active Aurora B (phospho-Thr232) from centromere/kinetochore region. Result is average from 3 replicate experiments \pm SD. 20 kinetochores were quantified from 5–10 cells in each experiment. HeLa cells were treated for 6 h with DMSO, 10 μ M VTT-006, 500 nM nocodazole, 100 nM Taxol or 5 μ M ZM447439 and 20 μ M MG132 followed by staining with anti-Aurora B, phospho-Thr232 and crest antibodies and DAPI. Aurora B (green), crest (red) and DAPI (blue) are shown in merge image. Star (*) denotes statistical significance, $p \le 0.05$. Scale bar = 10 μ m.





Supplementary Figure 6: Fine localization of Aurora B and active Aurora B (phospho-Thr232) in VTT-006 treated cells. (A) Merge images showing Aurora B (green) and crest (red) in cells treated with indicated compounds. (B) Close-up images of example kinetochore pairs selected from images in panel A. Images were acquired with a $100 \times$ objective using a confocal microscope. Line scans were prepared in Metamorph. For clarity, merge shows only Aurora B (green) and crest (red). HeLa cells were treated for 6 h with DMSO, 10 μ M VTT-006, 500 nM nocodazole, 100 nM Taxol or 5 μ M ZM447439 and 20 μ M MG132 followed by staining with anti-Aurora B, phospho-Thr232 and crest antibodies and DAPI. Scale bar = 10 μ m in A and 1 μ m in B.

Supplementary Movie 1: VTT-006 causes chromosome misalignment, mitotic arrest and aberrant cell division. See Supplementary Movie 1

Supplementary Movie 2: Control cells undergoing cell division. See Supplementary Movie 2

Supplementary Movie 3. MG132 treated cells remain arrested at metaphase. See Supplementary Movie 3

Supplementary Movie 4: VTT-006 disrupts metaphase plate alignment and causes chromosome movement towards spindle poles. See Supplementary Movie 4

Supplementary Movie 5: Taxol severely disrupts metaphase chromosome alignment. See Supplementary Movie 5

SUPPLEMENTARY MATERIALS AND METHODS

Cell culture

Cell line	Description	Growth medium	Source and year when cells were obtained
HeLa	cervical adenocarcinoma	DMEM (high glucose), 10% FBS, 20 mM HEPES, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, 2 mM L-glutamine, 0.1 mg/ml penicillin/ streptomycin	Prof. Gary Gorbsky, Oklahoma Medical Research Foundation, Oklahoma City, USA, 2004
HeLa-H2B- GFP	HeLa cells stably expressing H2B-GFP	DMEM (high glucose), 10% FBS, 20 mM HEPES, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, 2 mM L-glutamine, 0.1 mg/ml penicillin/ streptomycin, 2 µg/ml blasticidin	[57], 2006
GFP-Spc24- HeLa	HeLa cells stably expressing GFP-Spc24	DMEM (high glucose), 10% FBS, 20 mM HEPES, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, 2 mM L-glutamine, 0.1 mg/ml penicillin/ streptomycin, 200 µg/ml G418	Prof. Todd Stukenberg, University of Virginia, Charlottesville, USA, 2004
MCF10A	mammary epithelial cells	1:1 DMEM: HAM's F12, 5 % horse serum, 2 mM L-glutamine, 10 ug/ml insulin, 5 ug/ml hydrocortisone, 20 ng/ ml EGF, 100 ng/ml cholera toxin	ATCC CRL-10317, 2006
MCF7	breast adenocarcinoma	DMEM (low glucose), 10% FBS, 2 mM L-glutamine, 0.1 mg/ml penicillin/ streptomycin	ICLC HTL95021, 2004
MDA- MB-231	breast adenocarcinoma	DMEM (high glucose), 10% FBS, 0.1 mM non-essential amino acids, 2 mM L-glutamine, and 0.1 mg/ml penicillin/ streptomycin	ATCC HTB-26, 2004
MDA- MB-231 SA	breast adenocarcinoma	DMEM (high glucose), 10% FBS, 0.1 mM non-essential amino acids, 2 mM L-glutamine, and 0.1 mg/ml penicillin/ streptomycin	Prof. Theresa A. Guise, University of Virginia, Charlottesville, USA, 2004
LNCap	prostatic adenocarcinoma	RPMI 1640 supplemented with 10% FBS, 2 mM L-glutamine, and 0.1 mg/ml penicillin/streptomycin	ATCC CRL-1740, 2003
RWPE-1	prostate epithelial cells	KSFM containing BPE and EGF (Gibco 17005-042)	ATCC CRL-11609, 2006
RWPE-2-W99	tumorigenic derivative of RWPE-1	KSFM containing BPE and EGF (Gibco 17005-042)	ATCC CRL-2853, 2006
Ep156T	prostate epithelial cells	KSFM containing BPE and EGF (Gibco 17005-042)	Prof. Varda Rotter, Weitzmann Institute, Rehovot, Israel, 2006
HCT116	colorectal carcinoma	McCoy's 5A, 10% FBS, 2 mM L-glutamine, 0.1 mg/ml penicillin/ streptomycin	Prof. Lauri Aaltonen, University of Helsinki, Helsinki, Finland, 2009
A549	lung adenocarcinoma	RPMI 1640 supplemented with 10% FBS, 2 mM L-glutamine, and 0.1 mg/ml penicillin/streptomycin	ATCC CCL-185, 2005
Ovcar-3	ovary adenocarcinoma	RPMI 1640 supplemented with 10% FBS, 2 mM L-glutamine, and 0.1 mg/ml penicillin/streptomycin	[1], 2005

Statistics

Statistical analysis was performed with student's *t*-test.

SUPPLEMENTARY REFERENCE

 Hamilton TC, Young RC, McKoy WM, Grotzinger KR, Green JA, Whang-Peng J, Rogan AM, Ozols RF, Chu EW, Green WR. Characterization of a Human Ovarian Carcinoma Cell Line (NIH:OVCAR-3) with Androgen and Estrogen Receptors. Cancer Res. 1983; 43:5379–89. <u>PMID:6604576</u>