

Cancer cell repopulation after therapy: which is the mechanism?

Rewati Prakash¹ and Carlos M. Telleria^{1,2}

¹Experimental Pathology Unit, Department of Pathology, Faculty of Medicine and Health Sciences, McGill University, Montreal, QC, Canada

²Cancer Research Program, Research Institute, McGill University Health Centre, Montreal, QC, Canada

Correspondence to: Carlos M. Telleria, **email:** carlos.telleria@mcgill.ca

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ABSTRACT

Cancer cell repopulation after therapy is a phenomenon that leads to therapeutic failure with the consequent relapse of the disease. The process is understudied and mechanisms need to be uncovered. Here we discuss the issue of cancer cell repopulation after chemo- and radio-therapies. We compile evidence alleging that the repopulation of cancer cells can be originated from either cancer stem cells resistant to therapy, cancer cells that in response to therapy become polyploid and thereafter germinate into near-diploid rapid proliferating cells, and/or cells that respond to treatment undergoing senescence as a transient mechanism to survive, followed by the reinitiation of the cell cycle. Approaches targeted to prevent this post-therapy cancer cell repopulation should be uncovered to prevent tumor relapse and thus increase overall survival from this devastating disease.

INTRODUCTION

The past two decades have brought great progress in the treatment of cancer as patients with the disease live longer having access to better diagnosis and therapeutic approaches. However, the disease remains incurable. One of the reasons for the high resilience of this disease is that cancer cells hide and escape from therapies thus leading to cancer recurrence. The process whereby cells escape therapy is referred to as cancer cell repopulation [1] which is a phenomenon that has been mathematically modeled [2]. It was first thought that this was a biological mechanism limited to the tumor microenvironment whereby chemo- and radio-therapies were not efficiently distributed within the tumor to kill all cells with the capacity to propagate the disease [3]. However, the phenomenon can be recreated *in vitro*. For instance, we have shown that ovarian and non-small cell lung cancer cells highly sensitive to platinum drugs repopulate a culture despite the fact that supra-pharmacological doses of the chemotherapeutic agents were utilized in the experiments [4–6]; even though the majority of the cells were killed by the treatments, always few cells remain in culture with the acquired capability to recreate a similar population of cancer cells if provided with nutrients,

space, time, and not incurring in further insults. Likewise, cell repopulation has been demonstrated in breast and prostate cancers [7–9]. Nevertheless, despite being a well-documented phenomenon, the mechanisms involved in cancer cell repopulation remain poorly understood. In this perspective article we summarize three putative, not mutually exclusive molecular mechanisms that can drive the relapse of cancers via the regrowth of tumor cells that escape the initial treatment insult of chemotherapy and/or radiotherapy (Figure 1).

PUTATIVE MECHANISMS

Cancer stem cells

One of the first explanations for cancer cell repopulation is that the repopulating cells are derived from cancer stem cells. This theory implies that cancers are heterogeneous and have a progeny within which rapidly proliferating cells may be more easily killed by the therapy, sparing cells with stem-like phenotypes that are less differentiated and have cancer-initiating properties; they are termed cancer progenitor cells or cancer stem cells, which divide more slowly and therefore become spared by drugs and radiation. This residual therapy

resistant population is capable to regenerate the disease via transit-amplifying cells originating from a niche enriched in cancer stem cells [10, 11]. These cells become abundant after therapy likely because of the hypoxic conditions of the tumor microenvironment [12]. Cancer stem cells reside in hypoxic niches and use energy from glycolysis to gain protection from reactive oxygen species (ROS) usually generated during oxidative phosphorylation; they also divide asymmetrically to preserve their identity by preventing further mutations [13]. To demonstrate their complexity, it has been shown that in many cancer types they can interconvert within the tumor niche into non-cancer initiating cells via the epithelial-to-mesenchymal transition [14]. In addition, they have unique biomarkers and signaling pathways that vary among cancer types [15, 16]; these markers and pathways are essential to locate these scarce cells within a tumor. Whereas standard chemo- and radio-therapies seem to kill more differentiated cancer cells than cancer initiating cells [17], the ones with cancer stem cell properties develop resistance to treatment, persist within a niche, and regenerate a tumor by induction of proliferation upon escape from mitotic arrest [17].

It has been reported that the stem-like cells that resist treatment may also be influenced in a paracrine manner by dying cancer cells through a mechanism

initially reported in wound healing regenerative processes called “Phoenix rising”; this program involves cell death followed by compensatory proliferation driven by active caspase-3 leading to the generation of arachidonic acid as precursor of prostaglandin E2, which is needed for stem cell proliferation [9, 17, 18]. This phenomenon has been mathematically modeled [19] and is a putative mechanism whereby therapy-induced dying cells release factors that promote cancer cell repopulation.

A long-term yet challenging approach to abrogate the repopulating capacity of a tumor requires targeted approaches, which may not affect the bulk of the tumor, yet may eradicate the cancer propagating capacity of cancer stem cells only; targeting the slow dividing cancer initiating cells should lead to impaired recurrence [13, 16]. Finally, targeting pathways within the cancer stem cell niche or using immunotherapeutic strategies may provide a good opportunity to prevent cancer cell repopulation [20–22].

Polyloid giant cancer cells

Another feasible mechanism to justify cancer cell repopulation is that repopulating cells are derivatives of polyploid giant cancer cells formed in response to

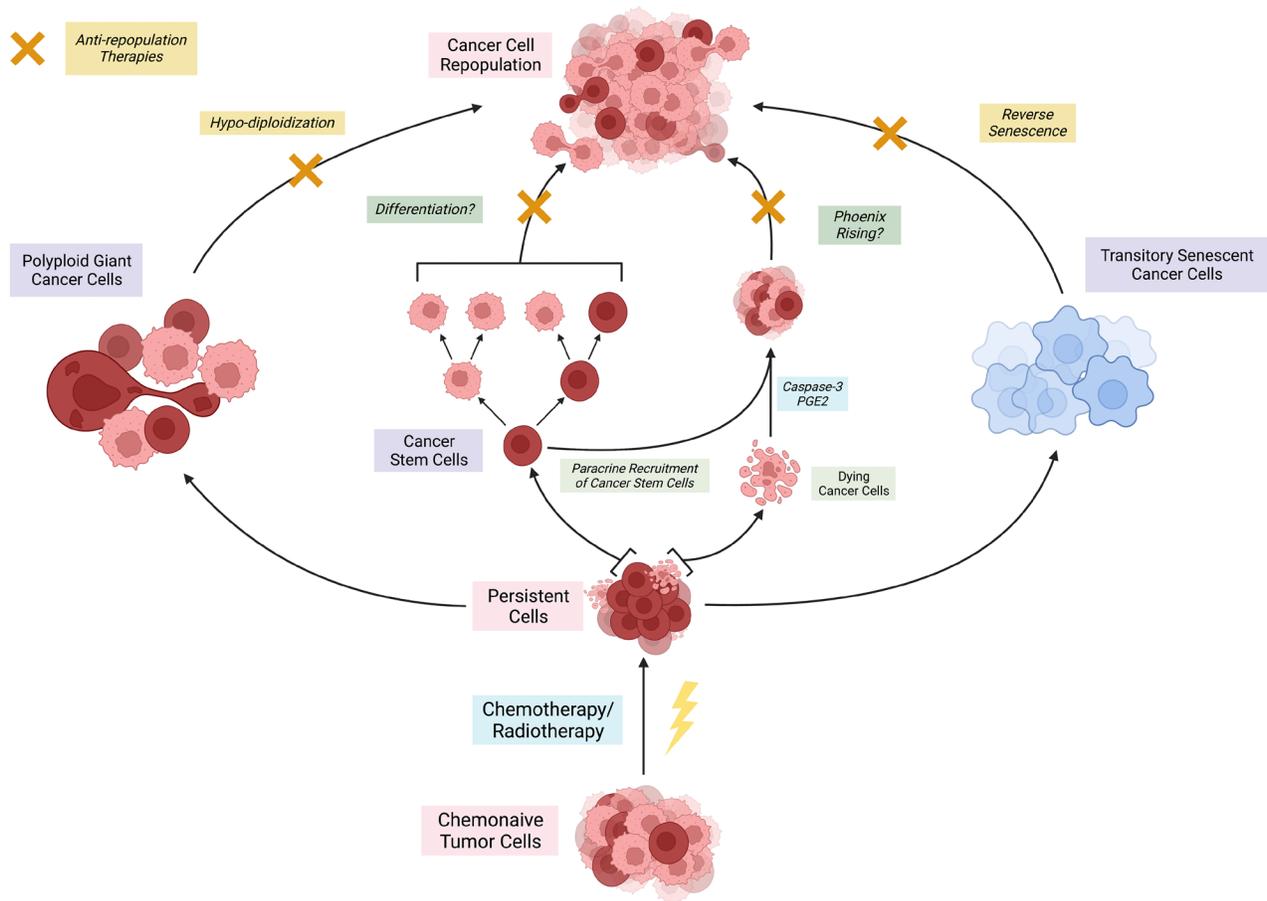


Figure 1: Presumed models of tumor cell repopulation after escaping chemoradiation.

therapies. These giant cells can be induced by hypoxia, chemotherapy, and radiation therapy [23–26]. However, with time, the polyploid giant cells are capable to give rise to smaller, near diploid highly proliferative cells via a mechanism coined as “neosis”, “hypo-diploidization”, or “reverse ploidy” [27, 28]. This is a modality of cell division generated in cells that escape mitotic crisis by undergoing budding followed by nuclear division and asymmetric cytokinesis resulting in the formation of aneuploid and mitotically active cells with genome stability [29, 30]. The polyploid giant cells are usually formed in response to the stress of chemoradiation, with the majority of them undergoing cell death. However, few survive isolating the chromosomes required by diploid tumor cells and producing a chemoresistant progeny via depolyploidization [31–33]. The polyploid giant cells arise as a consequence of endoreplication leading to the formation of mononucleated giant cells first, followed by cytokinetic failure and cell-cell fusion, thus giving rise to daughter cells able to proliferate for a long-term and carrying new chromosome alterations that cause distant metastases [32, 34, 35]. The emerging cells from polyploidy giant progenitors inherit stem-like properties [36] with capacity to differentiate into multiple malignant cell types [37]. The polyploid giant cells give rise to a near diploid progeny that gradually develops a more aggressive phenotype with passaging [38]. The overall process of hypo-diploidization utilizes meiosis-specific genes while cells eliminate excess chromatin [39, 40]. It is anticipated that uncovering the mechanism of reverse ploidy should guide researchers into developing targeted therapies that may prevent tumor recurrence.

Transient senescence cells

Growing evidence supports the idea that cancer cell repopulation happens as a consequence of transitory senescence. It is hypothesized that a rare percentage of cancer cells escape treatment by undergoing transient cell cycle arrest acquiring a senescence phenotype. This phenomenon has been often considered irreversible and characterized by cells having a flat morphology, expressing senescence-associated beta galactosidase due to enlargement of the lysosomal compartment, with formation of heterochromatic foci and endowed with a unique secretory program [41, 42]. One of the most important function of senescence is tumor suppression as these cells limit tumor progression by upregulating p53, p16, and p21, and are cleared by the immune system to limit tumorigenesis in usually premalignant lesions, or following cancer therapy [43, 44]. However, if senescence persists, it can also have detrimental effects in cancer tissues because the secretory phenotype of senescence cells generate a pro-inflammatory condition that favors tumor progression [42, 45].

The irreversibility of the senescence phenotype has nonetheless been heavily disputed. For instance

senescent cells with low expression of tumor suppressor p16^{ink4} resume cell growth upon inactivation of tumor suppressor p53 [46], p53 null lung cancer cells escape senescence induced by various drugs including cisplatin, camptothecin, etoposide, paclitaxel and vindesine by upregulating cyclin-dependent kinase 1 (Cdk1) [47], senescent colon and breast cancer cells regain proliferative capacity upon exposure to doxorubicin [48, 49], whereas senescent melanoma cells proliferate again upon the overexpression of the inhibitor of apoptosis protein survivin [50]. Escape from senescence has been also reported in breast cancer cell cultures exposed to conventional chemotherapy with the escaping cells expressing stem cell markers (high CD133 and Oct-4), low levels of ROS, and increased antioxidant enzymes [51]. Reinforcing the concept that indeed a senescence phenotype is not irreversible, a recent work, using a model of acute myeloid leukemia (AML) demonstrates that following chemotherapy, AML cells enter a senescence-like phenotype that repopulate the tumor leading to AML recurrence [52]. Of interest the phenomenon occurred by induction of embryonic diapause-like dormancy transcriptional signature and stemness reprogramming. It was also shown in this model that the induction of senescence, tumor survival and tumor persistence is dependent on ataxia telangiectasia and Rad3-related protein (ATR) involved in DNA damage/repair. In sum, there is mounting evidence that senescence is a transitional mechanism that can be induced by an array of therapies in various cancers and that is reversible with cells re-entering the cell cycle and repopulating a tumor. Consequently, it is questionable whether developing drugs to induce senescence as a manner to arrest cell growth in tumors is a clever way to stop cancer recurrence. Instead, usage of senolytic agents such as anti-BCL-2 family of proteins [42] or ATR inhibitors [52] may be a manner to eliminate transitory senescence cells. Perhaps the better approach to eliminate cancer cell repopulation is a combination treatment involving first chemoradiation-induced transitory senescence, followed by senolytic therapies as recently discussed by Wang and colleagues [42].

Author contributions

CMT researched the scientific literature. RP and CMT wrote the article. RP compiled the figure. Both authors approved the final version of the manuscript.

CONFLICTS OF INTEREST

Authors have no conflicts of interest to declare.

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REFERENCES

1. Kim JJ, Tannock IF. Repopulation of cancer cells during therapy: an important cause of treatment failure. *Nat Rev Cancer*. 2005; 5:516–25. <https://doi.org/10.1038/nrc1650>. PMID:15965493
2. Marcu L, Bezak E. Modelling of tumour repopulation after chemotherapy. *Australas Phys Eng Sci Med*. 2010; 33:265–70. <https://doi.org/10.1007/s13246-010-0026-4>. PMID:20652783
3. Davis AJ, Tannock JF. Repopulation of tumour cells between cycles of chemotherapy: a neglected factor. *Lancet Oncol*. 2000; 1:86–93. [https://doi.org/10.1016/s1470-2045\(00\)00019-x](https://doi.org/10.1016/s1470-2045(00)00019-x). PMID:11905673
4. Freeburg EM, Goyeneche AA, Telleria CM. Mifepristone abrogates repopulation of ovarian cancer cells in between courses of cisplatin treatment. *Int J Oncol*. 2009; 34:743–55. <https://doi.org/10.3892/ijo.00000200>. PMID:19212679
5. Gamarra-Luques CD, Goyeneche AA, Hapon MB, Telleria CM. Mifepristone prevents repopulation of ovarian cancer cells escaping cisplatin-paclitaxel therapy. *BMC Cancer*. 2012; 12:200. <https://doi.org/10.1186/1471-2407-12-200>. PMID:22642877
6. Kapperman HE, Goyeneche AA, Telleria CM. Mifepristone inhibits non-small cell lung carcinoma cellular escape from DNA damaging cisplatin. *Cancer Cell Int*. 2018; 18:185. <https://doi.org/10.1186/s12935-018-0683-z>. PMID:30479564
7. Wu L, Birlle DC, Tannock IF. Effects of the mammalian target of rapamycin inhibitor CCI-779 used alone or with chemotherapy on human prostate cancer cells and xenografts. *Cancer Res*. 2005; 65:2825–31. <https://doi.org/10.1158/0008-5472.CAN-04-3137>. PMID:15805283
8. Wu L, Tannock IF. Effect of the selective estrogen receptor modulator arzoxifen on repopulation of hormone-responsive breast cancer xenografts between courses of chemotherapy. *Clin Cancer Res*. 2005; 11:8195–200. <https://doi.org/10.1158/1078-0432.CCR-05-1258>. PMID:16299252
9. Corsi F, Capradossi F, Pelliccia A, Briganti S, Bruni E, Traversa E, Torino F, Reichle A, Ghibelli L. Apoptosis as Driver of Therapy-Induced Cancer Repopulation and Acquired Cell-Resistance (CRAC): A Simple In Vitro Model of Phoenix Rising in Prostate Cancer. *Int J Mol Sci*. 2022; 23:1152. <https://doi.org/10.3390/ijms23031152>. PMID:35163077
10. Malik B, Nie D. Cancer stem cells and resistance to chemo and radio therapy. *Front Biosci (Elite Ed)*. 2012; 4:2142–49. <https://doi.org/10.2741/531>. PMID:22202026
11. Neuzil J, Stantic M, Zobalova R, Chladova J, Wang X, Prochazka L, Dong L, Andera L, Ralph SJ. Tumour-initiating cells vs. cancer ‘stem’ cells and CD133: what’s in the name? *Biochem Biophys Res Commun*. 2007; 355:855–59. <https://doi.org/10.1016/j.bbrc.2007.01.159>. PMID:17307142
12. Heddleston JM, Li Z, McLendon RE, Hjelmeland AB, Rich JN. The hypoxic microenvironment maintains glioblastoma stem cells and promotes reprogramming towards a cancer stem cell phenotype. *Cell Cycle*. 2009; 8:3274–84. <https://doi.org/10.4161/cc.8.20.9701>. PMID:19770585
13. Battle E, Clevers H. Cancer stem cells revisited. *Nat Med*. 2017; 23:1124–34. <https://doi.org/10.1038/nm.4409>. PMID:28985214
14. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Brisken C, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell*. 2008; 133:704–15. <https://doi.org/10.1016/j.cell.2008.03.027>. PMID:18485877
15. Kuşoğlu A, Biray Avcı Ç. Cancer stem cells: A brief review of the current status. *Gene*. 2019; 681:80–85. <https://doi.org/10.1016/j.gene.2018.09.052>. PMID:30268439
16. Clevers H. The cancer stem cell: premises, promises and challenges. *Nat Med*. 2011; 17:313–19. <https://doi.org/10.1038/nm.2304>. PMID:21386835
17. Esmatabadi MJ, Bakhshinejad B, Motlagh FM, Babashah S, Sadeghizadeh M. Therapeutic resistance and cancer recurrence mechanisms: Unfolding the story of tumour coming back. *J Biosci*. 2016; 41:497–506. <https://doi.org/10.1007/s12038-016-9624-y>. PMID:27581940
18. Li F, Huang Q, Chen J, Peng Y, Roop DR, Bedford JS, Li CY. Apoptotic cells activate the “phoenix rising” pathway to promote wound healing and tissue regeneration. *Sci Signal*. 2010; 3:ra13. <https://doi.org/10.1126/scisignal.2000634>. PMID:20179271
19. Liu C, Li CY, Yuan F. Mathematical modeling of the Phoenix Rising pathway. *PLoS Comput Biol*. 2014; 10:e1003461. <https://doi.org/10.1371/journal.pcbi.1003461>. PMID:24516373
20. Anastas JN, Moon RT. WNT signalling pathways as therapeutic targets in cancer. *Nat Rev Cancer*. 2013; 13:11–26. <https://doi.org/10.1038/nrc3419>. PMID:23258168
21. Plaks V, Kong N, Werb Z. The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell*. 2015; 16:225–38. <https://doi.org/10.1016/j.stem.2015.02.015>. PMID:25748930
22. Atashzar MR, Baharlou R, Karami J, Abdollahi H, Rezaei R, Pourramezan F, Zoljalali Moghaddam SH. Cancer stem cells: A review from origin to therapeutic implications. *J Cell Physiol*. 2020; 235:790–803. <https://doi.org/10.1002/jcp.29044>. PMID:31286518
23. Fei F, Zhang M, Li B, Zhao L, Wang H, Liu L, Li Y, Ding P, Gu Y, Zhang X, Jiang T, Zhu S, Zhang S. Formation of Polyploid Giant Cancer Cells Involves in the Prognostic Value of Neoadjuvant Chemoradiation in Locally Advanced Rectal Cancer. *J Oncol*. 2019; 2019:2316436. <https://doi.org/10.1155/2019/2316436>. PMID:31558902
24. Chen J, Niu N, Zhang J, Qi L, Shen W, Donkena KV, Feng Z, Liu J. Polyploid Giant Cancer Cells (PGCCs): The Evil Roots of Cancer. *Curr Cancer Drug Targets*. 2019; 19:360–

67. <https://doi.org/10.2174/1568009618666180703154233>. PMID:29968537
25. Zhang Z, Feng X, Deng Z, Cheng J, Wang Y, Zhao M, Zhao Y, He S, Huang Q. Irradiation-induced polyploid giant cancer cells are involved in tumor cell repopulation via neosis. *Mol Oncol*. 2021; 15:2219–34. <https://doi.org/10.1002/1878-0261.12913>. PMID:33523579
 26. Niu N, Yao J, Bast RC, Sood AK, Liu J. IL-6 promotes drug resistance through formation of polyploid giant cancer cells and stromal fibroblast reprogramming. *Oncogenesis*. 2021; 10:65. <https://doi.org/10.1038/s41389-021-00349-4>. PMID:34588424
 27. Puig PE, Guilly MN, Bouchot A, Droin N, Cathelin D, Bouyer F, Favier L, Ghiringhelli F, Kroemer G, Solary E, Martin F, Chauffert B. Tumor cells can escape DNA-damaging cisplatin through DNA endoreduplication and reversible polyploidy. *Cell Biol Int*. 2008; 32:1031–43. <https://doi.org/10.1016/j.cellbi.2008.04.021>. PMID:18550395
 28. Sundaram M, Guernsey DL, Rajaraman MM, Rajaraman R. Neosis: a novel type of cell division in cancer. *Cancer Biol Ther*. 2004; 3:207–18. <https://doi.org/10.4161/cbt.3.2.663>. PMID:14726689
 29. Erenpreisa J, Kalejs M, Ianzini F, Kosmacek EA, Mackey MA, Emzinsh D, Cragg MS, Ivanov A, Illidge TM. Segregation of genomes in polyploid tumour cells following mitotic catastrophe. *Cell Biol Int*. 2005; 29:1005–11. <https://doi.org/10.1016/j.cellbi.2005.10.008>. PMID:16314119
 30. Saini G, Joshi S, Garlapati C, Li H, Kong J, Krishnamurthy J, Reid MD, Aneja R. Polyploid giant cancer cell characterization: New frontiers in predicting response to chemotherapy in breast cancer. *Semin Cancer Biol*. 2022; 81:220–31. <https://doi.org/10.1016/j.semcancer.2021.03.017>. PMID:33766651
 31. Song Y, Zhao Y, Deng Z, Zhao R, Huang Q. Stress-Induced Polyploid Giant Cancer Cells: Unique Way of Formation and Non-Negligible Characteristics. *Front Oncol*. 2021; 11:724781. <https://doi.org/10.3389/fonc.2021.724781>. PMID:34527590
 32. Niu N, Zhang J, Zhang N, Mercado-Urbe I, Tao F, Han Z, Pathak S, Multani AS, Kuang J, Yao J, Bast RC, Sood AK, Hung MC, Liu J. Linking genomic reorganization to tumor initiation via the giant cell cycle. *Oncogenesis*. 2016; 5:e281. <https://doi.org/10.1038/oncsis.2016.75>. PMID:27991913
 33. Niu N, Mercado-Urbe I, Liu J. Dedifferentiation into blastomere-like cancer stem cells via formation of polyploid giant cancer cells. *Oncogene*. 2017; 36:4887–900. <https://doi.org/10.1038/onc.2017.72>. PMID:28436947
 34. Moein S, Adibi R, da Silva Meirelles L, Nardi NB, Gheisari Y. Cancer regeneration: Polyploid cells are the key drivers of tumor progression. *Biochim Biophys Acta Rev Cancer*. 2020; 1874:188408. <https://doi.org/10.1016/j.bbcan.2020.188408>. PMID:32827584
 35. Øvrebø JI, Edgar BA. Polyploidy in tissue homeostasis and regeneration. *Development*. 2018; 145:dev156034. <https://doi.org/10.1242/dev.156034>. PMID:30021843
 36. Salmina K, Jankevics E, Huna A, Perminov D, Radovica I, Klymenko T, Ivanov A, Jascenko E, Scherthan H, Cragg M, Erenpreisa J. Up-regulation of the embryonic self-renewal network through reversible polyploidy in irradiated p53-mutant tumour cells. *Exp Cell Res*. 2010; 316:2099–112. <https://doi.org/10.1016/j.yexcr.2010.04.030>. PMID:20457152
 37. Díaz-Carballo D, Saka S, Klein J, Rennkamp T, Acikelli AH, Malak S, Jastrow H, Wennemuth G, Tempfer C, Schmitz I, Tannapfel A, Strumberg D. A Distinct Oncogenerative Multinucleated Cancer Cell Serves as a Source of Stemness and Tumor Heterogeneity. *Cancer Res*. 2018; 78:2318–31. <https://doi.org/10.1158/0008-5472.CAN-17-1861>. PMID:29440172
 38. Wang X, Zheng M, Fei F, Li C, Du J, Liu K, Li Y, Zhang S. EMT-related protein expression in polyploid giant cancer cells and their daughter cells with different passages after triptolide treatment. *Med Oncol*. 2019; 36:82. <https://doi.org/10.1007/s12032-019-1303-z>. PMID:31407170
 39. Ianzini F, Kosmacek EA, Nelson ES, Napoli E, Erenpreisa J, Kalejs M, Mackey MA. Activation of meiosis-specific genes is associated with depolyploidization of human tumor cells following radiation-induced mitotic catastrophe. *Cancer Res*. 2009; 69:2296–304. <https://doi.org/10.1158/0008-5472.CAN-08-3364>. PMID:19258501
 40. Kalejs M, Ivanov A, Plakhins G, Cragg MS, Emzinsh D, Illidge TM, Erenpreisa J. Upregulation of meiosis-specific genes in lymphoma cell lines following genotoxic insult and induction of mitotic catastrophe. *BMC Cancer*. 2006; 6:6. <https://doi.org/10.1186/1471-2407-6-6>. PMID:16401344
 41. Di Micco R, Krizhanovsky V, Baker D, d'Adda di Fagagna F. Cellular senescence in ageing: from mechanisms to therapeutic opportunities. *Nat Rev Mol Cell Biol*. 2021; 22:75–95. <https://doi.org/10.1038/s41580-020-00314-w>. PMID:33328614
 42. Wang L, Lankhorst L, Bernards R. Exploiting senescence for the treatment of cancer. *Nat Rev Cancer*. 2022; 22:340–55. <https://doi.org/10.1038/s41568-022-00450-9>. PMID:35241831
 43. Xue W, Zender L, Miething C, Dickins RA, Hernando E, Krizhanovsky V, Cordon-Cardo C, Lowe SW. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature*. 2007; 445:656–60. <https://doi.org/10.1038/nature05529>. PMID:17251933
 44. Kang TW, Yevsa T, Woller N, Hoenicke L, Wuestefeld T, Dauch D, Hohmeyer A, Gereke M, Rudalska R, Potapova A, Iken M, Vucur M, Weiss S, et al. Senescence surveillance of pre-malignant hepatocytes limits liver cancer development. *Nature*. 2011; 479:547–51. <https://doi.org/10.1038/nature10599>. PMID:22080947
 45. Schmitt CA, Wang B, Demaria M. Senescence and cancer - role and therapeutic opportunities. *Nat Rev Clin Oncol*. 2022; 19:619–36. <https://doi.org/10.1038/s41571-022-00668-4>. PMID:36045302
 46. Beauséjour CM, Krtolica A, Galimi F, Narita M, Lowe SW, Yaswen P, Campisi J. Reversal of human cellular

- senescence: roles of the p53 and p16 pathways. *EMBO J.* 2003; 22:4212–22. <https://doi.org/10.1093/emboj/cdg417>. PMID:12912919
47. Roberson RS, Kussick SJ, Vallieres E, Chen SY, Wu DY. Escape from therapy-induced accelerated cellular senescence in p53-null lung cancer cells and in human lung cancers. *Cancer Res.* 2005; 65:2795–803. <https://doi.org/10.1158/0008-5472.CAN-04-1270>. PMID:15805280
48. Sliwinska MA, Mosieniak G, Wolanin K, Babik A, Piwocka K, Magalska A, Szczepanowska J, Fronk J, Sikora E. Induction of senescence with doxorubicin leads to increased genomic instability of HCT116 cells. *Mech Ageing Dev.* 2009; 130:24–32. <https://doi.org/10.1016/j.mad.2008.04.011>. PMID:18538372
49. Elmore LW, Di X, Dumur C, Holt SE, Gewirtz DA. Evasion of a single-step, chemotherapy-induced senescence in breast cancer cells: implications for treatment response. *Clin Cancer Res.* 2005; 11:2637–43. <https://doi.org/10.1158/1078-0432.CCR-04-1462>. PMID:15814644
50. La Porta CA, Zapperi S, Sethna JP. Senescent cells in growing tumors: population dynamics and cancer stem cells. *PLoS Comput Biol.* 2012; 8:e1002316. <https://doi.org/10.1371/journal.pcbi.1002316>. PMID:22275856
51. Achuthan S, Santhoshkumar TR, Prabhakar J, Nair SA, Pillai MR. Drug-induced senescence generates chemoresistant stemlike cells with low reactive oxygen species. *J Biol Chem.* 2011; 286:37813–29. <https://doi.org/10.1074/jbc.M110.200675>. PMID:21878644
52. Duy C, Li M, Teater M, Meydan C, Garrett-Bakelman FE, Lee TC, Chin CR, Durmaz C, Kawabata KC, Dhimolea E, Mitsiades CS, Doehner H, D'Andrea RJ, et al. Chemotherapy Induces Senescence-Like Resilient Cells Capable of Initiating AML Recurrence. *Cancer Discov.* 2021; 11:1542–61. <https://doi.org/10.1158/2159-8290.CD-20-1375>. PMID:33500244